MYH9 is associated with nondiabetic end-stage renal disease in African Americans


As end-stage renal disease (ESRD) has a four times higher incidence in African Americans compared to European Americans, we hypothesized that susceptibility alleles for ESRD have a higher frequency in the West African than the European gene pool. We carried out a genome-wide admixture scan in 1,372 ESRD cases and 806 controls and found a highly significant association between excess African ancestry and nondiabetic ESRD (lod score = 5.70) but not diabetic ESRD (lod = 0.47) on chromosome 22q12. Each copy of the European ancestral allele conferred a relative risk of 0.50 (95% CI = 0.39–0.63) compared to African ancestry. Multiple common SNPs (allele frequencies ranging from 0.2 to 0.6) in the gene encoding nonmuscle myosin heavy chain type II isoform A (MYH9) were associated with two to four times greater risk of nondiabetic ESRD and accounted for a large proportion of the excess risk of ESRD observed in African compared to European Americans.

End-stage renal disease (ESRD) is the near-total loss of kidney function, requiring treatment of 472,000 individuals with dialysis or transplantation in the United States1. Diabetes and hypertension are the two leading reported causes of treated ESRD in the United States, accounting for 44% and 27% of incident cases, respectively1. African Americans have consistently had a much higher rate of ESRD than European Americans in the United States. In 2005, African Americans had a 3.7 times higher age-adjusted risk of ESRD. The risk ratio by ancestry. Multiple common SNPs (allele frequencies ranging from 0.2 to 0.6) in the gene encoding nonmuscle myosin heavy chain type II isoform A (MYH9) were associated with two to four times greater risk of nondiabetic ESRD and accounted for a large proportion of the excess risk of ESRD observed in African compared to European Americans.

End-stage renal disease (ESRD) is an excellent phenotype for genome-wide association analysis, an approach with enhanced power to detect common variants of kidney disease in African Americans even after these factors are taken into account. Family studies show clustering of ESRD independent of hypertension and diabetes5,6, with one large study showing stronger aggregation in African Americans8.

Studies attempting to detect susceptibility genes for ESRD and other complex diseases are challenging because of the late age of onset, causing difficulty in collecting multiply affected families, and because linkage analysis has suggested that there are no genes of high penetrance (fourfold increased risk) in populations of European descent, the focus of most published studies7,8. For these reasons, ESRD is an excellent phenotype for genome-wide association analysis, an approach with enhanced power to detect common variants of
modest penetration, and with the further advantage that unrelated
individuals can be studied.

We carried out a scan for ESRD risk loci using a particular type of
genome-wide association analysis termed admixture mapping or
mapping by admixture linkage disequilibrium (MALD)\(^9\)–\(^11\). Admixture
mapping is particularly suitable for finding genetic risk alleles that
differ in frequency between populations, which we hypothesized
might be the case for ESRD.

The basic principle of admixture mapping relies on the small
proportion of genetic variants that differ in frequency across populations
of different ancestries\(^12\). When mixing occurs between genetically
heterogeneous populations, the resulting admixed population inherits
chromosomal regions of either one ancestry or the other, and these
regions can be identified by genotyping markers that show substan-
tially different allele frequencies between ancestral populations.
Admixture mapping methodology is feasible as a result of the
development of a map of admixture mapping markers in African
Americans\(^13\) and appropriate analytical methods\(^14\),\(^15\). Moreover, the
admixture mapping methodology has been validated as a promising
way of finding susceptibility loci for common complex conditions,
such as prostate cancer\(^16\) and multiple sclerosis\(^17\).

The central hypothesis of this study was that some ESRD suscept-
ibility alleles are present at higher frequency in Africans than in
European Americans. Thus, the identification of ESRD susceptibility
alleles was possible by screening the genome in African Americans with
ESRD, searching for regions of the genome where individuals with the
disease have more (or less) African ancestry than their genome-wide
average. Not only have previous studies provided evidence for a genetic
basis to kidney disease and ESRD\(^7\),\(^18\), it has been hypothesized that
there may exist a common set of susceptibility genes for progression to
ESRD irrespective of the inciting cause, for example, diabetic or
nondiabetic\(^19\). A secondary goal of our study was to test the hypothesis
that the genetic variants that confer a higher risk for ESRD in African
Americans are relevant to a broad range of ESRD phenotypes.

RESULTS

Initial admixture scan results using all ESRD cases

A total of 1,372 ESRD cases and 806 controls without the presence of
either an elevated serum creatinine concentration or albuminuria at
recruitment were included in the initial scan using 1,354 markers. The
mean age of ESRD cases and non-ESRD controls was 53 (s.d. = 13)
and 46 (s.d. = 12) years, respectively. Fifty-three percent of cases were
male, compared to 39% of controls.

The mean age of dialysis initiation in the 841 cases was 48 years
(s.d. = 13), and these individuals spent an average of 3 years on
dialysis (information was not available for all participants). Approx-
imately half of the ESRD cases (n = 703; 51%) had diabetes (75%) with
known types classified as type 2 diabetes) as the etiology of ESRD, and
669 had nondiabetic causes. The mean diabetes duration was 21 years
(s.d. = 9.6) with a mean age of onset of 36 years (s.d. = 13) among the
591 cases with diabetic ESRD. Most of the participants with nondia-
betic ESRD had hypertension reported as the cause of ESRD (n = 347;
25% of all ESRD), 87 had focal segmental glomerulosclerosis (FSGS),
69 had HIV-related nephropathy, 126 had other glomerulonephritis
from primary or systemic diseases and 40 had other causes. The
mean duration of hypertension was 13 years among 266 hypertensive
ESRD cases.

We carried out the initial scan of 1,372 ESRD cases and 806
controls with 1,354 ancestry informative markers using 12 pre-
specified European ancestry risk models ranging from 0.3 to 1.5, in
increments of 0.1, and a control risk of 1. The genome-wide lod
score, which was averaged across all risk models to account for
multiple comparison, showed a suggestive association with ESRD
on chromosome 22 (genome-wide score of 1.67, where values
greater than 2 are considered genome-wide significant; Table 1).
In the region of rs4417800 on chromosome 22, the single-point lod
score was 4.63 (highest of all chromosomes), and the highest-
scored risk model was of one copy of the European ancestral allele,
conferring a 0.7-fold risk of ESRD compared to the equivalent with
African ancestry (reference group).

Significant admixture-generated signal for nondiabetic ESRD

To determine whether susceptibility genes for diabetic ESRD and
nondiabetic ESRD were different, we carried out admixture scans
limited to either diabetic (n = 703) or nondiabetic ESRD (n = 669)
as an a priori subgroup analysis. The same group of 806 non-ESRD
controls was used for both scans. We did not observe any genome-wide
significant association in the diabetic ESRD scan (genome-wide score
of 0.47; Table 1), with the strongest locus-specific signal arising from
chromosome 11 (lod = 2.79, which does not meet the threshold of
locus-specific lod > 5 for evidence of association; Table 2 and Fig. 1a).
In contrast, a highly significant genome-wide score of 5.70 was
observed for the nondiabetic ESRD subset. The highest single-point
lod score (8.56) was detected on chromosome 22 for the nondiabetic
ESRD analysis (Table 2 and Fig. 1a). The risk model with the strongest
score corresponded to one copy of the European ancestral allele,
conferring a risk of 0.50 (95% CI = 0.39–0.63) compared to African
ancestry (reference group). Upon further narrowing of the risk models
(ranging from 0.4 to 0.6 in increments of 0.02), we obtained a
genome-wide score of 9.31 in this region. In addition, the estimated
European ancestry in this region on chromosome 22 was lower than
the average across the entire genome among nondiabetic ESRD cases
but not controls (Fig. 1b). The case for the strong difference between
the diabetic and the nondiabetic ESRD cohorts was further strength-
ened by a case-control analysis in which the individuals with non-
diabetic ESRD were used as cases and the diabetic ESRD cases were
used as controls at the locus on chromosome 22. This analysis also
showed a highly significant increase in African ancestry in nondiabetic
ESRD compared to diabetic ESRD cases at the chromosome 22 peak
(T statistic = −10.6, P = 2.94 × 10\(^{-15}\)).

To assess the robustness of the chromosome 22 peak, we further
evaluated whether the evidence for association was dependent on a
particular marker. We divided the data into even- and odd-numbered
SNPs. Each set of SNPs independently showed evidence of association

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**Table 1 Summary of results from admixture scans in African Americans with end-stage renal disease (ESRD)**

<table>
<thead>
<tr>
<th>ESRD phenotype</th>
<th>SNPs</th>
<th>Cases/Controls</th>
<th>Genome score</th>
<th>Lod score at chr. 22 peak</th>
</tr>
</thead>
<tbody>
<tr>
<td>All ESRD</td>
<td>1,354</td>
<td>1,372/806</td>
<td>1.67</td>
<td>4.55</td>
</tr>
<tr>
<td>DM</td>
<td>1,351</td>
<td>703/806</td>
<td>0.47</td>
<td>−0.52</td>
</tr>
<tr>
<td>Non-DM</td>
<td>1,354</td>
<td>669/806</td>
<td>5.70(^a)</td>
<td>8.56</td>
</tr>
<tr>
<td>HTN</td>
<td>1,352</td>
<td>347/806</td>
<td>−0.16</td>
<td>1.79</td>
</tr>
<tr>
<td>FSGS</td>
<td>1,350</td>
<td>87/806</td>
<td>0.10</td>
<td>2.47</td>
</tr>
<tr>
<td>GN</td>
<td>1,351</td>
<td>126/806</td>
<td>0.01</td>
<td>1.75</td>
</tr>
<tr>
<td>HIV</td>
<td>1,348</td>
<td>69/806</td>
<td>0.11</td>
<td>2.09</td>
</tr>
</tbody>
</table>

ESRD, end stage renal disease; DM, diabetic; Non-DM, nondiabetic; FSGS, focal segmental glomerulosclerosis; HTN, hypertensive; GN, glomerulonephritis; HIV, human immunodeficiency virus nephropathy. Non-DM includes HTN, FSGS, HIV, GN and 40 cases with other causes.

\(^a\)Genome score is the genome-wide lod score which corrects for multiple comparison by averaging locus-specific lod scores across the entire genome; a score above 1.0 is considered significant association and a score above 1.0 is considered suggestive association.

T. O. A. V. O. M. E. A. R. S. 4. 0
A study of FSGS in African Americans. This related study also

tions between authors of the present study and those of a concurrent

Fig. 2

Mb (that an approximate 95% CI for the region was 31.388650–35.039798

where the lod score was within 0.86 of its maximum, we estimated

nondiabetic ESRD in African Americans. By identifying the region

underlying genetic variant(s) contributing to the increased risk for

ESRD cases, we calculated a confidence interval for the position of the

Given the consistency of a signal across different groups of nondiabetic

glomerulonephritis (FSGS) ESRD. Although the risk models were not iden-

tical for different ESRD types, with the strongest effect observed for

FSGS, the signals of association were consistent across all ESRD types,

with the strongest association signals generated in the vicinity of

rs5999762 from the entire admixture SNP panel. The evidence of

association was present for HIV nephropathy but much weaker

with the admixture signal was present for HIV-related ESRD but not controls.

(CEU) from Utah and the African-ancestry samples from Yoruba

(YRI) from the International HapMap Project and found to be

significantly associated with FSGS and hypertensive ESRD in the

related study. All 14 MYH9 SNPs were highly significantly associated

with nondiabetic ESRD using either the conservative general genetic

model parameterizing three genotype-specific risks (Supplementary

Tables 1 and 2 online) or a recessive genetic model (P values as low as

3.63 × 10^{-15} after correcting for global ancestry; Table 3) at a

Bonferroni-corrected α level of 0.003 (0.05/14). As anticipated, none

of the SNPs were associated with diabetic ESRD (lowest P value = 0.03

for rs16996677 after adjustment for global ancestry; Table 3). The

Segmentation

Analysis of MYH9 on chromosome 22q12

Given the consistency of a signal across different groups of nondiabetic

ESRD cases, we calculated a confidence interval for the position of the

underlying genetic variant(s) contributing to the increased risk for

nondiabetic ESRD in African Americans. By identifying the region

where the lod score was within 0.86 of its maximum, we estimated

that an approximate 95% CI for the region was 31.388650–35.039798

Mb (Fig. 2). The region contains 22 genes (17 known). Although

many are expressed in the kidney, MYH9, the gene encoding the

nonmuscle myosin heavy chain 9, is the most highly expressed

compared to the other 16 known genes in this region. Moreover, we

chose to study MYH9 as a biological positional candidate gene on

the basis of the existing knowledge of the importance of MYH9

mutations in a number of kidney diseases and ongoing collabora-

tions between authors of the present study and those of a concurrent

study of FSGS in African Americans. This related study also

identified a region of interest on chromosome 22 and further

established MYH9 as a risk factor for FSGS and subsequent hyperten-

sive ESRD. We genotyped 14 SNPs in MYH9 known to have high

allele frequency differences between the European-ancestry samples

(genome-wide score = 6.32 for even-numbered SNP scan and 5.57 for

odd-numbered SNP scan).

To assess whether the significant signal from the nondiabetic ESRD

scan was generated by one particular subtype of ESRD, we carried out

additional admixture scans for the major subtypes of nondiabetic

ESRD: hypertensive ESRD and FSGS (case numbers for glomerulone-

phritis and HIV-related ESRD were smaller). At the same locus on

chromosome 22q (rs5999762), the best risk models, as indicated by

relative risks, and the corresponding locus-specific lod scores were as

follows: 0.5 (lod = 8.467) for all nondiabetic, 0.8 (lod = 1.70) for

hypertensive, and 0.35 (lod = 2.48) for focal and segmental glomer-

ulosclerosis (FSGS) ESRD. Although the risk models were not identi-

tical for different ESRD types, with the strongest effect observed for

FSGS, the signals of association were consistent across all ESRD types,

with the strongest association signals generated in the vicinity of

rs5999762 from the entire admixture SNP panel. The evidence of

admixture signal was present for HIV nephropathy but much weaker

for secondary glomerulonephritis (Table 1).

Figure 1 Summary of admixture scan results. (a) Lod score from genome-wide scan for diabetic (gray) and nondiabetic (black) ESRD. The genome-wide score was 5.70 for nondiabetic ESRD and 0.47 for diabetic ESRD. The highest locus-specific lod for diabetic ESRD was 2.79 on chromosome 11 and 8.56 on chromosome 22 for nondiabetic ESRD. (b) Difference between estimated European ancestry at different loci of the genome and the genome-wide average for 669 nondiabetic ESRD (black) and 806 controls (gray). The estimated European ancestry on chromosome 22 was lower than the average across the entire genome among nondiabetic ESRD cases but not controls.

Figure 2 Region of association with nondiabetic ESRD on chromosome 22. The peak lod score was 9.31, and the 95% CI spanned from 31.388650 to 35.039798 Mb, covering 22 genes. We considered a lod score for association at a particular locus of >5 as approximately genome-wide significant.
Table 2 Summary of genome-wide scan results among African Americans with ESRD by subtypes (highest lod score by chromosome)

<table>
<thead>
<tr>
<th>Chr.</th>
<th>DM</th>
<th>Non-DM</th>
<th>HTN</th>
<th>FSGS</th>
<th>GN</th>
<th>HIV</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>0.64</td>
<td>1.5</td>
<td>0.95</td>
<td>0.16</td>
<td>0.39</td>
<td>0.8</td>
</tr>
<tr>
<td>2</td>
<td>1.7</td>
<td>1.45</td>
<td>0.86</td>
<td>0.4</td>
<td>1</td>
<td>0.56</td>
</tr>
<tr>
<td>3</td>
<td>-0.16</td>
<td>-0.37</td>
<td>-0.03</td>
<td>0.39</td>
<td>1.45</td>
<td>0.28</td>
</tr>
<tr>
<td>4</td>
<td>0.47</td>
<td>0.25</td>
<td>-0.11</td>
<td>0.26</td>
<td>0.37</td>
<td>0.14</td>
</tr>
<tr>
<td>5</td>
<td>0.31</td>
<td>-0.26</td>
<td>-0.17</td>
<td>0.45</td>
<td>0.16</td>
<td>0.52</td>
</tr>
<tr>
<td>6</td>
<td>0.55</td>
<td>2.41</td>
<td>1.23</td>
<td>0.58</td>
<td>0.4</td>
<td>0.39</td>
</tr>
<tr>
<td>7</td>
<td>-0.07</td>
<td>-0.25</td>
<td>0.69</td>
<td>0.9</td>
<td>0.91</td>
<td>0.29</td>
</tr>
<tr>
<td>8</td>
<td>1.09</td>
<td>0.75</td>
<td>1.03</td>
<td>0.21</td>
<td>0.67</td>
<td>0.53</td>
</tr>
<tr>
<td>9</td>
<td>-0.28</td>
<td>0.14</td>
<td>-0.17</td>
<td>0.1</td>
<td>0.42</td>
<td>0.23</td>
</tr>
<tr>
<td>10</td>
<td>0.12</td>
<td>1.49</td>
<td>0.81</td>
<td>0.32</td>
<td>0.61</td>
<td>0.72</td>
</tr>
<tr>
<td>11</td>
<td>2.79</td>
<td>-0.37</td>
<td>0.76</td>
<td>0.03</td>
<td>0.4</td>
<td>0.06</td>
</tr>
<tr>
<td>12</td>
<td>1.17</td>
<td>0.97</td>
<td>-0.05</td>
<td>0.37</td>
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<td>0.31</td>
</tr>
<tr>
<td>13</td>
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<td>0.3</td>
<td>0.61</td>
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<td>1.34</td>
</tr>
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<tr>
<td>15</td>
<td>1.06</td>
<td>-0.28</td>
<td>0.33</td>
<td>0.41</td>
<td>-0.15</td>
<td>0.25</td>
</tr>
<tr>
<td>16</td>
<td>0.34</td>
<td>-0.53</td>
<td>-0.23</td>
<td>0.38</td>
<td>0.01</td>
<td>0.47</td>
</tr>
<tr>
<td>17</td>
<td>0.37</td>
<td>0.18</td>
<td>0.3</td>
<td>0.1</td>
<td>0.8</td>
<td>0.35</td>
</tr>
<tr>
<td>18</td>
<td>0.41</td>
<td>0.03</td>
<td>-0.28</td>
<td>0.57</td>
<td>0.06</td>
<td>0.42</td>
</tr>
<tr>
<td>19</td>
<td>-0.23</td>
<td>0.46</td>
<td>-0.3</td>
<td>0.54</td>
<td>0.41</td>
<td>0.01</td>
</tr>
<tr>
<td>20</td>
<td>0.5</td>
<td>-0.28</td>
<td>-0.36</td>
<td>0.18</td>
<td>0.08</td>
<td>0.92</td>
</tr>
<tr>
<td>21</td>
<td>0.68</td>
<td>0.15</td>
<td>0.67</td>
<td>0.1</td>
<td>0.62</td>
<td>0.01</td>
</tr>
<tr>
<td>22</td>
<td>-0.52</td>
<td>8.56</td>
<td>1.79</td>
<td>2.47</td>
<td>1.75</td>
<td>2.09</td>
</tr>
<tr>
<td>X</td>
<td>0.82</td>
<td>0.32</td>
<td>0.15</td>
<td>0.11</td>
<td>0.74</td>
<td>0.94</td>
</tr>
</tbody>
</table>

DM, diabetic; Non-DM, nondiabetic; HTN, hypertensive; FSGS, focal segmental glomerulosclerosis; GN, glomerulonephritis; HIV, human immunodeficiency virus nephropathy.

Table 3 Adjusted odds ratio and 95% confidence interval of nondiabetic and diabetic ESRD for 14 MYH9 SNPs

<table>
<thead>
<tr>
<th>SNP</th>
<th>Position</th>
<th>Alleles</th>
<th>MAF (%)</th>
<th>Ref. allele</th>
<th>OR (95% CI)</th>
<th>P value, global</th>
<th>P value, global and local</th>
<th>OR 95% CI</th>
<th>P value, global</th>
<th>P value, global and local</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs7078</td>
<td>35007860</td>
<td>G/A</td>
<td>15.3</td>
<td>G</td>
<td>1.97 (1.52,2.55)</td>
<td>3.41 × 10⁻⁷</td>
<td>1.58 × 10⁻⁵</td>
<td>1.28 (1.01,1.62)</td>
<td>0.04</td>
<td>0.13</td>
</tr>
<tr>
<td>rs12107</td>
<td>35007928</td>
<td>A/G</td>
<td>9.3</td>
<td>A</td>
<td>1.57 (1.16,2.12)</td>
<td>3.64 × 10⁻³</td>
<td>0.01</td>
<td>0.90 (0.69,1.18)</td>
<td>0.45</td>
<td>0.76</td>
</tr>
<tr>
<td>rs375853</td>
<td>35009161</td>
<td>G/C</td>
<td>10.2</td>
<td>G</td>
<td>1.90 (1.38,2.62)</td>
<td>9.05 × 10⁻⁵</td>
<td>0.14</td>
<td>0.91 (0.69,1.19)</td>
<td>0.47</td>
<td>0.77</td>
</tr>
<tr>
<td>rs5756129</td>
<td>35014038</td>
<td>C/T</td>
<td>19.3</td>
<td>C</td>
<td>1.71 (1.35,2.17)</td>
<td>7.92 × 10⁻⁶</td>
<td>3.21 × 10⁻⁵</td>
<td>1.06 (0.85,1.32)</td>
<td>0.63</td>
<td>0.89</td>
</tr>
<tr>
<td>rs5756130</td>
<td>35014277</td>
<td>T/C</td>
<td>12.5</td>
<td>T</td>
<td>1.58 (1.22,2.08)</td>
<td>1.20 × 10⁻³</td>
<td>1.00 (0.78,1.28)</td>
<td>0.99</td>
<td>0.99</td>
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</tr>
<tr>
<td>rs875725</td>
<td>35021637</td>
<td>T/C</td>
<td>6.0</td>
<td>C</td>
<td>1.17 (0.84,1.65)</td>
<td>0.35</td>
<td>0.29</td>
<td>0.91 (0.66,1.25)</td>
<td>0.55</td>
<td>0.84</td>
</tr>
<tr>
<td>rs4821480</td>
<td>35025193</td>
<td>T/G</td>
<td>29.5</td>
<td>T</td>
<td>2.18 (1.73,2.73)</td>
<td>1.72 × 10⁻¹¹</td>
<td>1.36 × 10⁻⁶</td>
<td>0.95 (0.75,1.19)</td>
<td>0.65</td>
<td>0.90</td>
</tr>
<tr>
<td>rs2032487</td>
<td>35025374</td>
<td>T/C</td>
<td>33.9</td>
<td>T</td>
<td>2.41 (1.94,3.01)</td>
<td>4.51 × 10⁻₁⁰</td>
<td>2.78 × 10⁻⁸</td>
<td>1.11 (0.89,1.38)</td>
<td>0.35</td>
<td>0.64</td>
</tr>
<tr>
<td>rs4821481</td>
<td>35025888</td>
<td>T/C</td>
<td>34.7</td>
<td>T</td>
<td>2.40 (1.93,3.29)</td>
<td>3.63 × 10⁻¹⁰</td>
<td>2.80 × 10⁻⁸</td>
<td>1.07 (0.86,1.33)</td>
<td>0.53</td>
<td>0.82</td>
</tr>
<tr>
<td>rs3752462</td>
<td>35040129</td>
<td>C/T</td>
<td>25.6</td>
<td>C</td>
<td>2.11 (1.69,2.63)</td>
<td>5.19 × 10⁻¹¹</td>
<td>1.98 × 10⁻⁵</td>
<td>1.08 (0.87,1.33)</td>
<td>0.47</td>
<td>0.77</td>
</tr>
<tr>
<td>rs5756152</td>
<td>35042418</td>
<td>A/G</td>
<td>28.7</td>
<td>G</td>
<td>2.59 (1.88,3.35)</td>
<td>4.49 × 10⁻⁹</td>
<td>2.25 × 10⁻⁶</td>
<td>1.28 (0.91,1.83)</td>
<td>0.16</td>
<td>0.38</td>
</tr>
<tr>
<td>rs1005570</td>
<td>35045220</td>
<td>A/G</td>
<td>44.9</td>
<td>G</td>
<td>2.45 (1.93,3.12)</td>
<td>2.56 × 10⁻¹³</td>
<td>4.33 × 10⁻⁹</td>
<td>1.27 (0.98,1.63)</td>
<td>0.07</td>
<td>0.19</td>
</tr>
<tr>
<td>rs16996674</td>
<td>35056598</td>
<td>T/C</td>
<td>23.4</td>
<td>C</td>
<td>3.10 (2.15,4.47)</td>
<td>1.49 × 10⁻⁹</td>
<td>4.14 × 10⁻⁷</td>
<td>1.51 (1.01,2.27)</td>
<td>0.04</td>
<td>0.13</td>
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<td>rs16996677</td>
<td>35057229</td>
<td>A/G</td>
<td>26.8</td>
<td>G</td>
<td>3.03 (2.16,4.25)</td>
<td>1.61 × 10⁻¹⁰</td>
<td>1.47 × 10⁻⁷</td>
<td>1.50 (1.03,2.17)</td>
<td>0.03</td>
<td>0.10</td>
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</tbody>
</table>

OR and P value adjusted for global ancestry; P value adjusted for both global and local ancestry is also shown. MAF assumed by the recessive genetic model is the minor allele frequency in African Americans and refers to the allele listed first in the ‘Alleles’ column. Ref. allele refers to allele used as the reference group in the logistic regression.
Table 4 Adjusted odds ratio and 95% confidence interval of hypertensive and focal segmental glomerulosclerosis ESRD for 14 MYH9 SNPs

<table>
<thead>
<tr>
<th>SNP</th>
<th>Position</th>
<th>Alleles</th>
<th>MAF (%)</th>
<th>Ref. allele</th>
<th>OR (95% CI)</th>
<th>P value, global</th>
<th>P value, global and local</th>
<th>OR (95% CI)</th>
<th>P value, global</th>
<th>P value, global and local</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs7078</td>
<td>35007960</td>
<td>G/A</td>
<td>15.3</td>
<td>G</td>
<td>1.70 (1.24, 2.33)</td>
<td>1.04 × 10⁻³</td>
<td>4.03 × 10⁻³</td>
<td>3.52 (1.73, 16.7)</td>
<td>5.17 × 10⁻⁴</td>
<td>1.85 × 10⁻³</td>
</tr>
<tr>
<td>rs12107</td>
<td>35007928</td>
<td>A/G</td>
<td>09.3</td>
<td>A</td>
<td>1.34 (0.93, 1.93)</td>
<td>0.12</td>
<td>0.16</td>
<td>2.95 (1.26, 6.91)</td>
<td>0.01</td>
<td>0.02</td>
</tr>
<tr>
<td>rs738583</td>
<td>35009161</td>
<td>G/C</td>
<td>10.2</td>
<td>G</td>
<td>2.08 (1.36, 3.16)</td>
<td>6.79 × 10⁻⁴</td>
<td>0.02</td>
<td>1.79 (0.86, 3.7)</td>
<td>0.12</td>
<td>0.82</td>
</tr>
<tr>
<td>rs5756129</td>
<td>35014038</td>
<td>C/T</td>
<td>19.3</td>
<td>C</td>
<td>1.62 (1.21, 2.18)</td>
<td>1.19 × 10⁻³</td>
<td>2.04 × 10⁻³</td>
<td>2.81 (1.55, 5.07)</td>
<td>6.35 × 10⁻⁴</td>
<td>1.19 × 10⁻³</td>
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<tr>
<td>rs5756130</td>
<td>35014277</td>
<td>T/C</td>
<td>12.5</td>
<td>T</td>
<td>1.69 (1.19, 2.4)</td>
<td>3.45 × 10⁻³</td>
<td>3.36 × 10⁻³</td>
<td>2.42 (1.19, 4.93)</td>
<td>0.01</td>
<td>0.02</td>
</tr>
<tr>
<td>rs875725</td>
<td>3501637</td>
<td>T/C</td>
<td>06.0</td>
<td>C</td>
<td>1.01 (0.67, 1.51)</td>
<td>0.97</td>
<td>0.90</td>
<td>3.69 (1.14, 11.94)</td>
<td>0.03</td>
<td>0.03</td>
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<tr>
<td>rs4821480</td>
<td>35025193</td>
<td>T/G</td>
<td>29.5</td>
<td>T</td>
<td>2.07 (1.56, 2.74)</td>
<td>3.7 × 10⁻⁷</td>
<td>3.18 × 10⁻⁵</td>
<td>3.66 (1.16, 3.34)</td>
<td>3.98 × 10⁻⁶</td>
<td>4.54 × 10⁻³</td>
</tr>
<tr>
<td>rs2032487</td>
<td>35025374</td>
<td>T/C</td>
<td>33.9</td>
<td>T</td>
<td>2.28 (1.74, 2.98)</td>
<td>2.38 × 10⁻⁹</td>
<td>5.77 × 10⁻⁷</td>
<td>4.85 (2.82, 8.33)</td>
<td>1.09 × 10⁻⁸</td>
<td>6.14 × 10⁻⁵</td>
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<tr>
<td>rs4821481</td>
<td>35025888</td>
<td>T/C</td>
<td>34.7</td>
<td>T</td>
<td>2.32 (1.73, 3.03)</td>
<td>7.81 × 10⁻⁷</td>
<td>4.37 × 10⁻⁷</td>
<td>4.34 (2.61, 7.21)</td>
<td>1.45 × 10⁻⁸</td>
<td>9.57 × 10⁻⁵</td>
</tr>
<tr>
<td>rs3752462</td>
<td>35040129</td>
<td>C/T</td>
<td>25.6</td>
<td>C</td>
<td>2.00 (1.52, 2.64)</td>
<td>6.79 × 10⁻⁷</td>
<td>9.52 × 10⁻⁵</td>
<td>3.50 (2.01, 6.07)</td>
<td>8.65 × 10⁻⁶</td>
<td>2.53 × 10⁻³</td>
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<td>G</td>
<td>2.54 (1.76, 3.68)</td>
<td>7.71 × 10⁻⁷</td>
<td>1.59 × 10⁻⁵</td>
<td>4.63 (2.73, 8.78)</td>
<td>1.45 × 10⁻⁸</td>
<td>2.44 × 10⁻⁵</td>
</tr>
<tr>
<td>rs1005570</td>
<td>35045220</td>
<td>A/G</td>
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<td>G</td>
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<td>G</td>
<td>2.78 (1.81, 4.27)</td>
<td>2.83 × 10⁻⁶</td>
<td>3.56 × 10⁻⁵</td>
<td>6.84 (3.91, 11.97)</td>
<td>1.69 × 10⁻¹¹</td>
<td>6.82 × 10⁻⁹</td>
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OR and P value adjusted for global ancestry; P value adjusted for both global and local ancestry is also shown. MAF assumed by the recessive genetic model is the minor allele frequency in African Americans and refers to the allele listed first in the ‘Alleles’ column. Ref. allele refers to allele used as the reference group in the logistic regression.

Association with nondiabetic ESRD after control for ancestry

As expected, not only did MYH9 SNPs account for the admixture signal on chromosome 22, but they were significantly associated with nondiabetic ESRD even after accounting for difference in global ancestry. To understand the contribution of alleles to the phenotype16, we estimated the OR associated with each SNP for nondiabetic ESRD. For example, the OR associated with rs875725 for the A allele of rs4821481 had a frequency of 75.8% in YRI but only 6.1% in CEU. These three SNPs represent highly unusual differences (above the 99% percentile of autosomal SNPs in terms of allele frequency differentiation). Thus, the strong correlation of the allele with ancestry explains the strength of the admixture signal.

Notably, further upstream of rs2032487 were two other common SNPs (rs16996674 with a frequency of 23.4% for the high-risk allele T and rs16996677 with a frequency of 26.8% for the high-risk allele A) in intron 3 that were in almost perfect LD with each other and that were even more significantly associated with nondiabetic ESRD after adjustment for global and local ancestry. Assuming a three-genotypic risk model, those with either the AG or AA genotype at rs16996674 were about 1.63 (95% CI = 1.29–2.05) and 3.82 (95% CI = 2.67–5.47) (Table 4). Sixty-four percent of all nondiabetic ESRD cases had the CC genotype compared to 42% of controls.

Figure 3 Effect of MYH9 SNPs on the association between local ancestry in the region of chromosome 22q12 and nondiabetic ESRD. The results of the local ancestry estimates were obtained from 14 different ANCESTRYMAP runs, each containing one of the 14 SNPs at a time. The light gray bars indicate the highly significant associations between local ancestry at the locus of interest on chromosome 22q12 and case-control status. The dark gray bars show the greatly reduced associations between local ancestry and case-control status after the inclusion of MYH9 SNP in the logistic regression model. Three SNPs (rs4821480, rs2032487 and rs4821481), in particular, accounted for almost the entire significance of the association between local ancestry and case-control status. The LD pattern of 14 SNPs in MYH9 shown is that of the African-American FIND and CHOICE participants.
more likely to have nondiabetic ESRD than their counterparts with the GG genotype (Supplementary Table 1). The frequency of the AA and AG genotypes were 19% and 43%, respectively, in nondiabetic ESRD cases but only 7% and 39% in controls. The excess risk of nondiabetic ESRD associated with the A allele was also observed using a recessive model, where individuals with the AA genotype were about 3.03 times more likely to have nondiabetic ESRD than their counterparts who had either the GG or AG genotype (95% CI = 2.16–4.25; \( P = 1.61 \times 10^{-10} \)).

Table 3.

Upon cross tabulation of rs2032487 and rs16996677, both SNPs remained independently associated with nondiabetic ESRD. Although the two SNPs are correlated, they are not in complete LD \( (r^2 \sim 0.88) \), suggesting that these two SNPs are capturing a haplotype that contains the causal variant. Lastly, we performed both adjusted and stratified analyses to account for the effect of ancestry on rs2032487 and rs16996677. Both SNPs remained highly significantly associated with nondiabetic ESRD even after adjustment for both global and local ancestry (see Table 4 for ORs adjusted for only global ancestry and Supplementary Table 2 for ORs adjusted for both global and local ancestry). In analyses that were stratified by ancestral background of this region of the chromosome (zero, one or two copies of the African ancestral chromosome at this region), both SNPs remained associated with nondiabetic ESRD on the background of either one or two copies of the African ancestral chromosome at this region (ORs were not estimated for those with zero copies of the African ancestral chromosome at this region because only about 1% of the population had zero copies of the African ancestral chromosome at this region and about 84% of the population had two copies of the African ancestral chromosome at this region; Supplementary Table 5 online).

**DISCUSSION**

In this genome-wide admixture scan of 1,372 ESRD cases and 806 controls, we identified a locus on chromosome 22q12 that showed a significant excess of African ancestry among nondiabetic ESRD cases compared to controls without nephropathy. Moreover, we showed that multiple SNPs in MYH9 can account for the admixture-generated signal in this region. Although the evidence for MYH9 harboring one or more susceptibility alleles for nondiabetic ESRD in African Americans is exceedingly strong, it is not clear that the genetic variant responsible (the causal variant) has yet been identified, as none of the SNPs typed in the present study are known to influence either transcription or translation directly. Moreover, multiple MYH9 SNPs, including rs2032487 and rs16996677, were correlated with each other and were highly significantly associated with nondiabetic ESRD. Thus, it is likely that our results have simply identified genetic variants that are in strong linkage disequilibrium with the causal variant.

An intriguing result of the admixture scan is that, independent of the knowledge of the causal SNP, the results allow us to estimate how much of the epidemiologically increased risk for nondiabetic ESRD in African Americans as compared with European Americans may be accounted for by this locus. Using estimates and the assumptions made in the admixture scan, we calculate that compared with individuals who have entirely African ancestry at this locus on chromosome 22, individuals with one European chromosome at the locus are estimated to have 0.50 times the prevalence (95% CI = 0.39–0.63), and individuals with two European chromosomes have 0.5 \( \times \) 0.5 = 0.25 the prevalence. We estimate from this that if it were possible to reduce the prevalence of nondiabetic ESRD in African Americans (assuming 16.9% European ancestry on average) to the rate that would be expected if all the African Americans had inherited European ancestry at the locus, the prevalence of the nondiabetic ESRD in African Americans would decrease to 30% (95% CI = 19–45%) of its current estimate (for details of this calculation, see Supplementary Table 6 online).

The lack of association between MYH9 SNPs and diabetic ESRD is unlikely the result of low statistical power. With 703 diabetic ESRD cases and 806 controls, this study would have had close to 100% power to detect an OR of 2.4 (the OR associated with both rs2032487 and rs4821481, which has an \( r^2 \sim 1 \) with rs4821480) assuming that the frequency of the at-risk genotype is 56% in controls (percent of individuals who are homozygous for the at-risk allele at either SNP) and an \( \alpha \) of 0.003. The marked contrast in results for diabetic versus nondiabetic ESRD suggests that mutations in MYH9 are not strongly associated with diabetic ESRD, especially ESRD associated with type 2 diabetes. In diabetic ESRD, unlike other types of ESRD, the initiation of renal damage is due to hyperglycemia with an increase in glomerular filtration rate and mesangial proliferation. The inexorable progression of renal disease, once renal damage occurs, has led to the hypothesis that there are common underlying pathogenic pathways leading to ESRD. Although the lack of association of MYH9 in this study with diabetic kidney disease does not disprove the hypothesis that there are common mechanisms for progression of all forms of chronic kidney disease, it shows that there are at least some mechanisms of ESRD progression that are unique to specific diseases. In particular, the results indicate that MYH9 is important for the progression of nondiabetic ESRD but not for that of diabetic ESRD, although the present study was not able to test whether MYH9 may have differential effects on progression to ESRD depending on the type of diabetes (type 1 versus type 2). Thus, genetic variation in MYH9 may only be important in the kidney’s response to injury not caused by hyperglycemia.

MYH9 encodes the protein nonmuscle myosin heavy chain, class II, and isoform type A in eukaryotic cells. The gene is approximately 110 kb with 41 exons and is highly conserved among a number of mammalian species and very similar to other nonmuscle myosin isoforms. The protein is abundantly expressed in the kidney, liver and platelets. These proteins have a variety of cellular functions, such as in cellular polarity, architecture and trafficking. Rare mutations in MYH9 are associated with several mendelian conditions, including an autosomal dominant form of deafness (MIM160775), Epstein syndrome (MIM153560), Fechtner syndrome (MIM153640), May–Hegglin anomaly (MIM155100) and Sebastian syndrome (MIM153640); moreover, many of these conditions may include renal disease (Fig. 3).

Within the kidney, MYH9 expression occurs in the glomerulus, specifically the podocyte, peritubular capillaries and tubules. Aggregation of abnormal myosin and damage to the cytoskeleton of the podocyte and tubular cells could lead to progressive kidney disease; however, it remains to be determined how sequence variations in MYH9 directly result in development and progression of nondiabetic kidney disease.

In summary, common genetic variations in the MYH9 locus on chromosome 22q can account for much of the excess risk of nondiabetic ESRD observed in African Americans compared to European Americans, although environmental risk factors and interactions between genes and environment undoubtedly play an additional role as well. Moreover, there is also a distinct genetic susceptibility to nondiabetic ESRD among African Americans that is not evident among cases with diabetic ESRD, suggesting that the mechanisms leading from the initiation of chronic kidney disease to progression to ESRD may differ depending on the inciting cause. Thus, drug therapies that are often targeted at common pathways to
prevent kidney disease progression may need more specificity for effective treatment.

Finally, the identification of susceptibility alleles that exist in different frequencies between African Americans and European Americans has potential implications for future screening strategies for African Americans with nondiabetic kidney disease. Our finding suggests that many African American individuals reported as having hypertensive ESRD may actually have an identifiable etiology other than hypertension. This may explain why strict blood pressure control often fails to slow the progression of purported hypertensive kidney disease in many African Americans.

In summary, these findings suggest a previously unknown MYH9 pathway for kidney disease progression with major potential implications. Experiments should identify the causal variants, elucidate the pathophysiologic mechanisms, and evaluate strategies to influence this pathway that may decrease risk of progression to ESRD among African Americans. Further research can ultimately lead to the identification of high-risk individuals and the development of new approaches to combat the epidemic rise of ESRD in the United States and other nations.

METHODS

Subjects. Participants were recruited as part of the Family Investigation of Nephropathy and Diabetes (FINDD) and the Choices for Healthy Outcomes In Caring for ESRD (CHOICE) study. The FINDD study populations and design have been described in detail elsewhere. Briefly, participants were recruited from 11 participating centers. The African American MALD study recruited both diabetic and nondiabetic ESRD cases and spouse or partner controls. Subjects were recruited and samples collected according to the Declaration of Helsinki principles, and a certificate of confidentiality was filed at the National Institutes of Health. The CHOICE study is a national prospective cohort study of 1,041 ESRD participants (294 were African American) from 81 dialysis clinics associated with Dialysis Clinic, Inc., New Haven CAPD and the Hospital of St. Raphael that has been described previously. The study was approved by the institutional review boards at Johns Hopkins University and the associated clinical sites, and participants provided written informed consent.

From both studies, a total of 1,427 ESRD cases (883 from FINDD African American MALD study; 299 from FIND Family study; 245 from the CHOICE study) and 839 controls (only from the African American FIND MALD study) were included. Of these 2,256 cases and controls, 36 individuals were dropped during routine data check in ANCESTRYMAP. 17 were dropped for having potential gender inconsistencies between self-reported gender and probability of heterozygosity states on markers on the X chromosome, and 25 were dropped for having low genotype calling rates. Diabetes (primarily type 2 diabetes) was defined as self-reported and/or prevalent treatment with insulin and/or oral hypoglycemic agents or documented in the medical history. Diabetes, hypertension and kidney disease duration were obtained from the medical history and by medical record review. Information on specific type of diabetes obtained from medical chart abstraction was available on 160 of the diabetic ESRD cases: 118 (74%) of them had type 2 diabetes, 22 had type 1 diabetes (14%) and 20 (12%) had unknown type diabetes listed on their medical records. Only cases recruited from the Johns Hopkins FIND site had medical chart abstraction review to classify diabetes. Age of onset for diabetes, hypertension and kidney disease was by self report. ESRD due to diabetic nephropathy was defined by having two of the following three clinical criteria: (i) onset of diabetes ≥ 5 years before renal replacement therapy; (ii) documented diabetic retinopathy; or (iii) proteinuria (≥ 3.0 g protein/g creatinine). ESRD due to nondiabetic nephropathy was defined as biopsy-proven nondiabetic kidney disease or by cause of ESRD abstracted from medical chart charts obtained during physician visits and hospitalizations. ESRD from hypertension was defined by one of the following: (i) evidence of hypertensive nephropathy based on previous biopsy findings, or (ii) the exclusion of all other causes of ESRD by the participants’ primary nephrologist. In addition, in those cases of hypertensive ESRD without a biopsy, less than 6% had evidence of significant proteinuria of > 3.0 g protein/g creatinine at the time of presentation. We also specifically excluded all congenital, known monogenic forms and cancer-related renal diseases.

Genotyping. We attempted genotyping of 1,536 SNPs using the Illumina BeadLab platform for the initial admixture mapping scan. Details of the genotyping for this experiment are as described previously. We constructed this panel mainly by using the panel of ancestry informative markers from a previously published map and then improving the panel by mining new ancestry informative markers from the SNP datasets supplemented with > 1,500 new markers identified by Hinds et al. and the Phase 2 International Haplotype Map, and validating them to confirm that they were indeed ancestry informative. These SNPs were then prioritized as most informative about West African versus European ancestry, according to their predicted usefulness for determining ancestry. After quality checks, including the requirement that at least 85% of SNPs were successfully genotyped for each sample and that none of the SNPs were in linkage disequilibrium in the ancestral populations, 182 SNPs were excluded from analysis (call rates for each of the SNPs kept are shown in Supplementary Table 7 online). The physical genome positions used in this study are based on build 33 of the public genome reference. To obtain the genetic positions, we used the Rutgers Integrated Map as previously described. Using the TaqMan pre-designed genotyping assays (Applied Biosystems) under standard conditions, we genotyped an additional 14 SNPs in MYH9, selected for having a pronounced frequency difference between reference African (YRI) and European (CEU) populations.

Admixture scan. We used ANCESTRYMAP to perform admixture mapping analyses. This program uses a Markov chain Monte Carlo–based (MCMC) methodology. To accumulate evidence of association in these models, we averaged the Bayes factors emerging from each model at each point in the genome, taking the log10 of this number to produce a lod score. We considered a lod score for association at a particular locus of > 5 as approximately genome-wide significant. To obtain a formal assessment of statistical significance on a genome-wide level, we calculated an additional statistic that averaged the risks specified in the models as genome-wide Bayes factors and took the log10: a value > 2 indicates statistically significant association to the phenotype. For the initial scan, we ran ANCESTRYMAP for a burn-in period of 100 iterations with 200 follow-on iterations and averaged scores obtained from 12 pre-specified European ancestry risk models ranging from 0.3 to 1.5 in increments of 0.1. For example, the model testing a risk of 1.5 assumed a 1.5-fold increased risk due to inheritance of one copy of European ancestral allele for cases, with a control risk of 1. To calculate the 95% CI for the position of the disease locus once we found an association, we first summarized the evidence for association by taking the sum of the likelihood ratios across the entire locus. Then, starting at the peak of the locus, we moved in both directions until the region included 95% of the value of the sum.

To test for association between SNPs and ESRD above and beyond the confounding effect of ancestry association, we obtained local and global estimates of ancestry by using ANCESTRYMAP. Global ancestry was obtained using all of the markers from the initial scan. We obtained the estimate of local ancestry on MYH9 14 times, each time using one of the MYH9 SNPs along with the other SNPs from the original scan. Estimates obtained from these 14 SNPs were used to adjust for the effect of ancestry locally on the association between MYH9 SNPs and case-control status definition.

Association between SNPs in MYH9 and various case-control status definitions was determined by the means of ORs, 95% confidence intervals and P values obtained from logistic regression models (SAS v10). We assumed a general model parameterizing three genotypic risks for initial association analyses (Supplementary Tables 1 and 2). On the basis of ORs derived from the general models, we further tested and presented the results for the recessive model. A Bonferroni-corrected Ũ of 0.003 (0.05/14 SNPs) was used to declare significance. The logistic regression was used to perform a case-control comparison to determine whether MYH9 SNPs were associated with diabetic ESRD (as opposed to nondiabetic ESRD). The Breslow-Day test for heterogeneity was used to determine whether the associations between MYH9 SNPs and ESRD status differed significantly by diabetes status.
Note: Supplementary information is available on the Nature Genetics website.

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